



# Pathophysiology of COVID-19

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BOLTZMANN  
INSTITUTE  
Traumatology

The Research Center in Cooperation with AUVA





# Disclosures

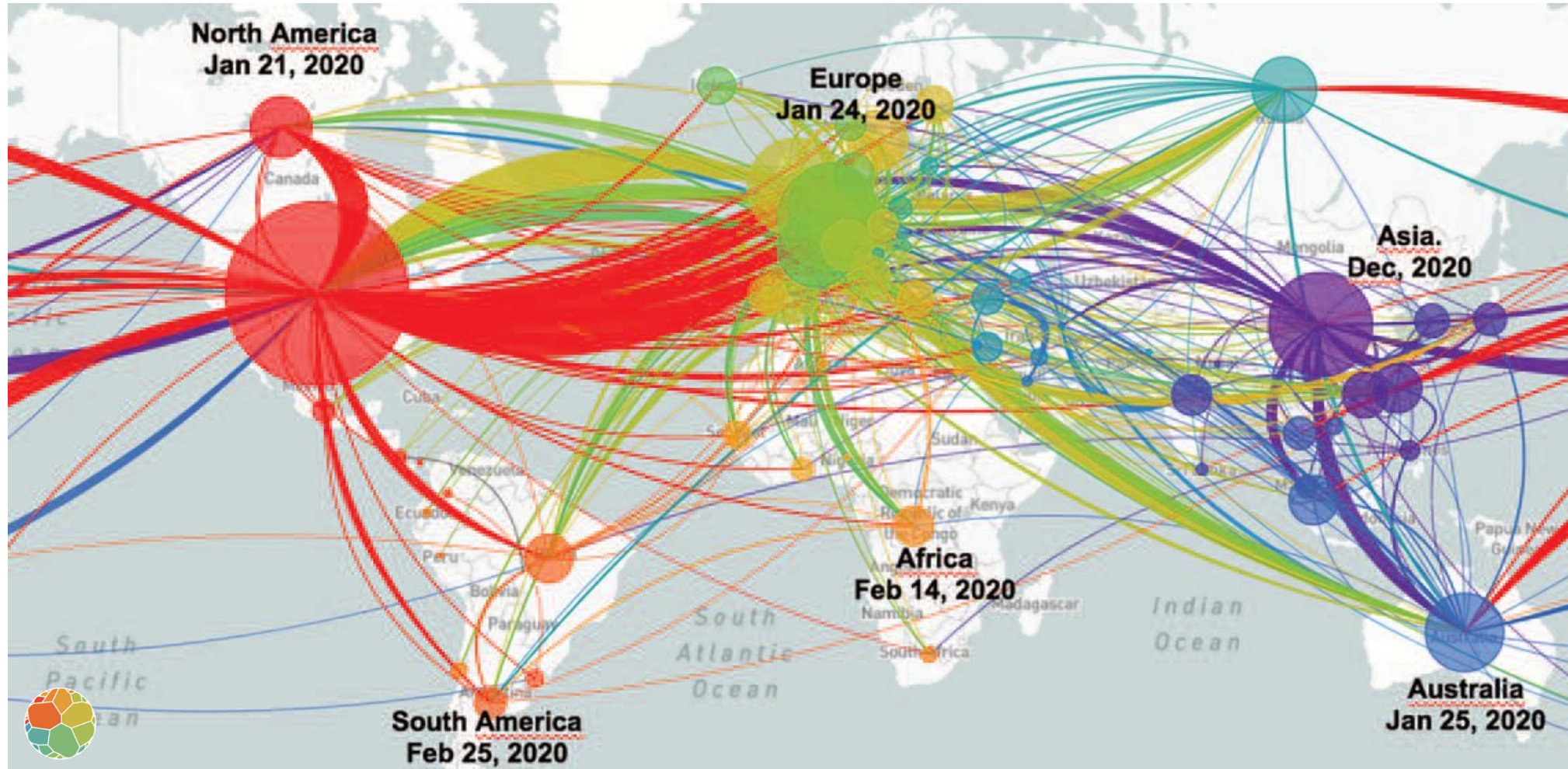
I do not have conflict of interest for this talk

I have never personally treated any COVID-19 patients

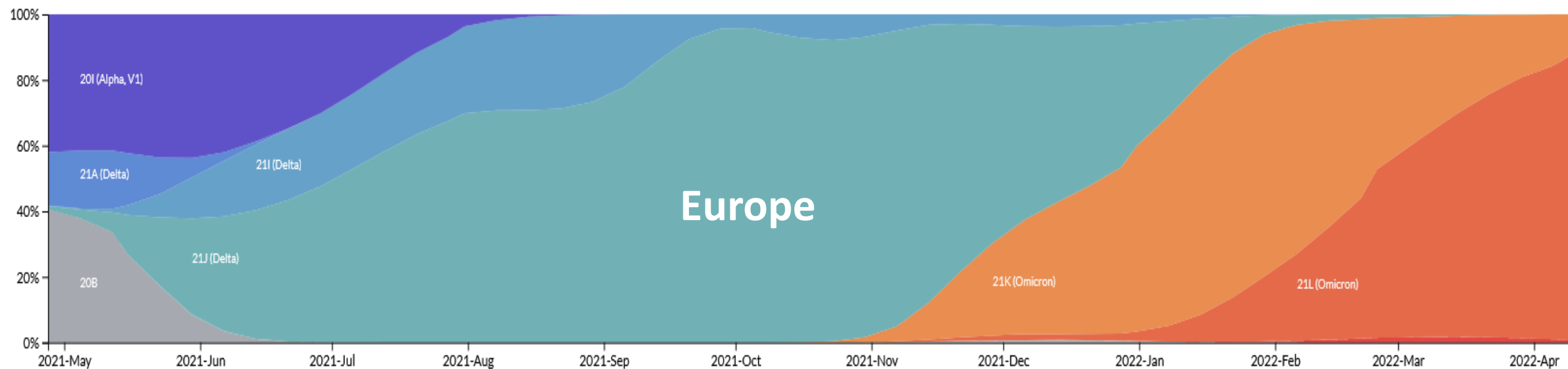
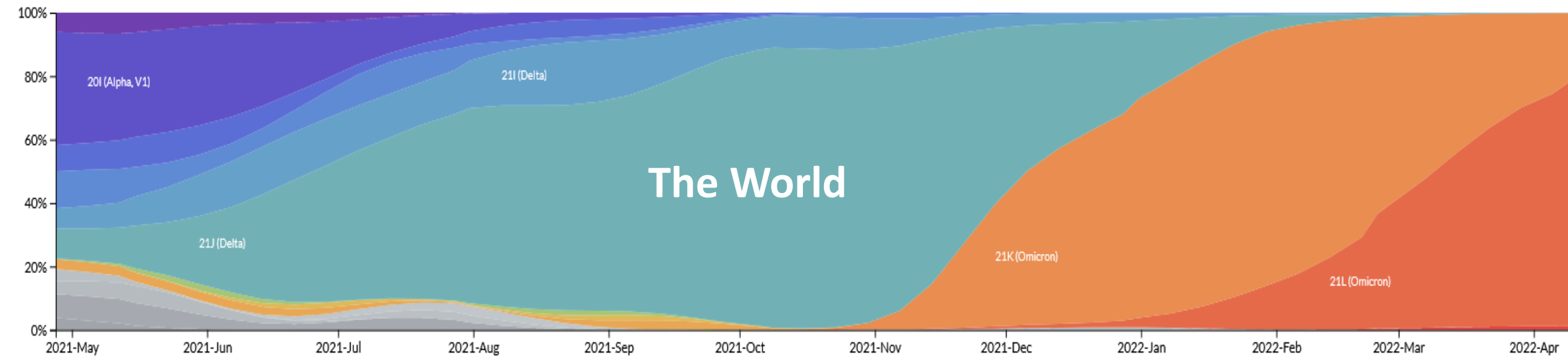
The Shock & Lancet Resp Med review interpretations of C-19 data presented here are a team effort as they largely stem from the collegial brainstorming of ESS and EGIS groups that created them.



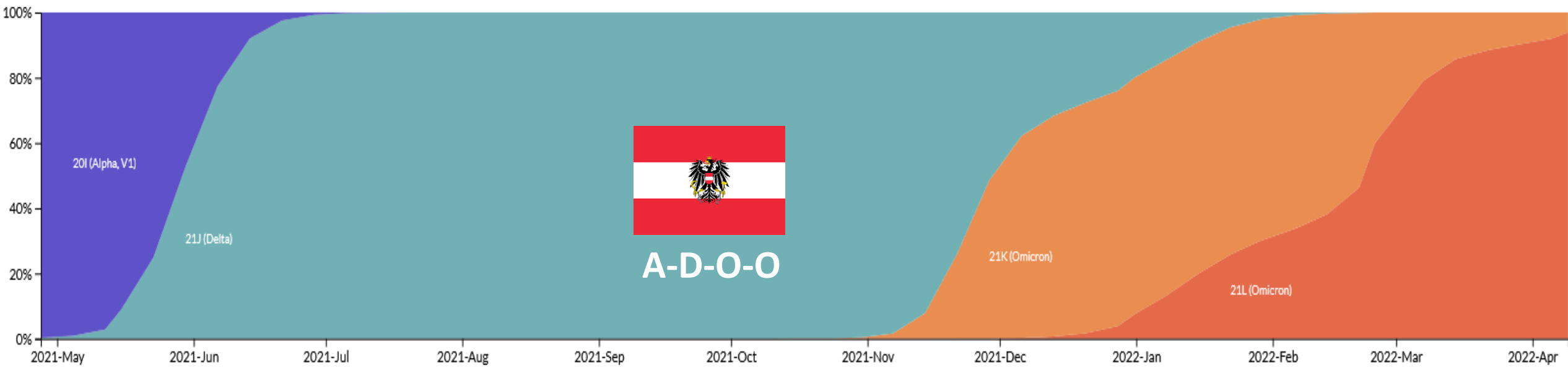
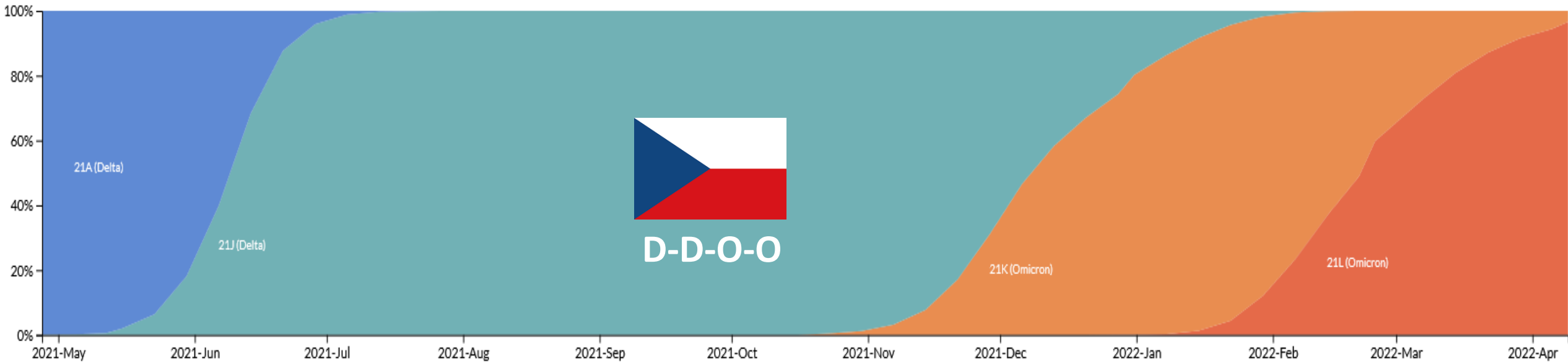
# COVID-19: the Beginning...



# SARS-CoV-2: the Evolution of the Variants



# SARS-CoV-2: the Evolution of the Variants



# Virus Tropism Mainly Correlates with ACE2 Tissue Distribution

## Organ/cell tropism of SARS-CoV-2



### Lungs

- ✓ Basal, ciliated and club cells
- ✓ AT2 cells
- ✓ Proliferative KRT7<sup>+</sup> epithelial cells
- ✓ Vascular endothelial cells



### Trachea

- ✓ Ciliated and goblet cells of the mucosa
- ✓ Epithelial cells of the conduits and the glands



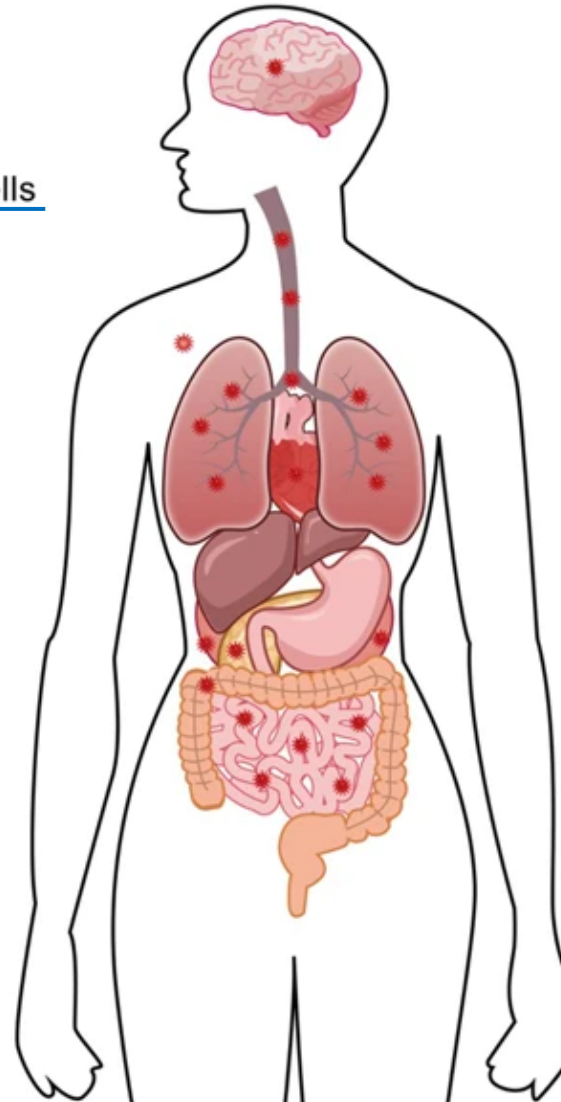
### Intestines

- ✓ Epithelial cells in the mucosa and the glands



### Skin

- ✓ Secretory luminal cells in the sweat glands
- ✓ Vascular endothelial cells



### Kidneys

- ✓ Epithelial cells in the distal tubule and collecting ducts
- ✓ Vascular endothelial cells



### Pancreas

- ✓ Epithelial cells in the langerhans, glands, and intra-islet ducts
- ✓ Vascular endothelial cells



### Brain

- ✓ Vascular endothelial cells



### Heart

- ✓ Vascular endothelial cells



### SARS-CoV-2



# Schematic Diagram of SARS-CoV-2 Entry Pathways

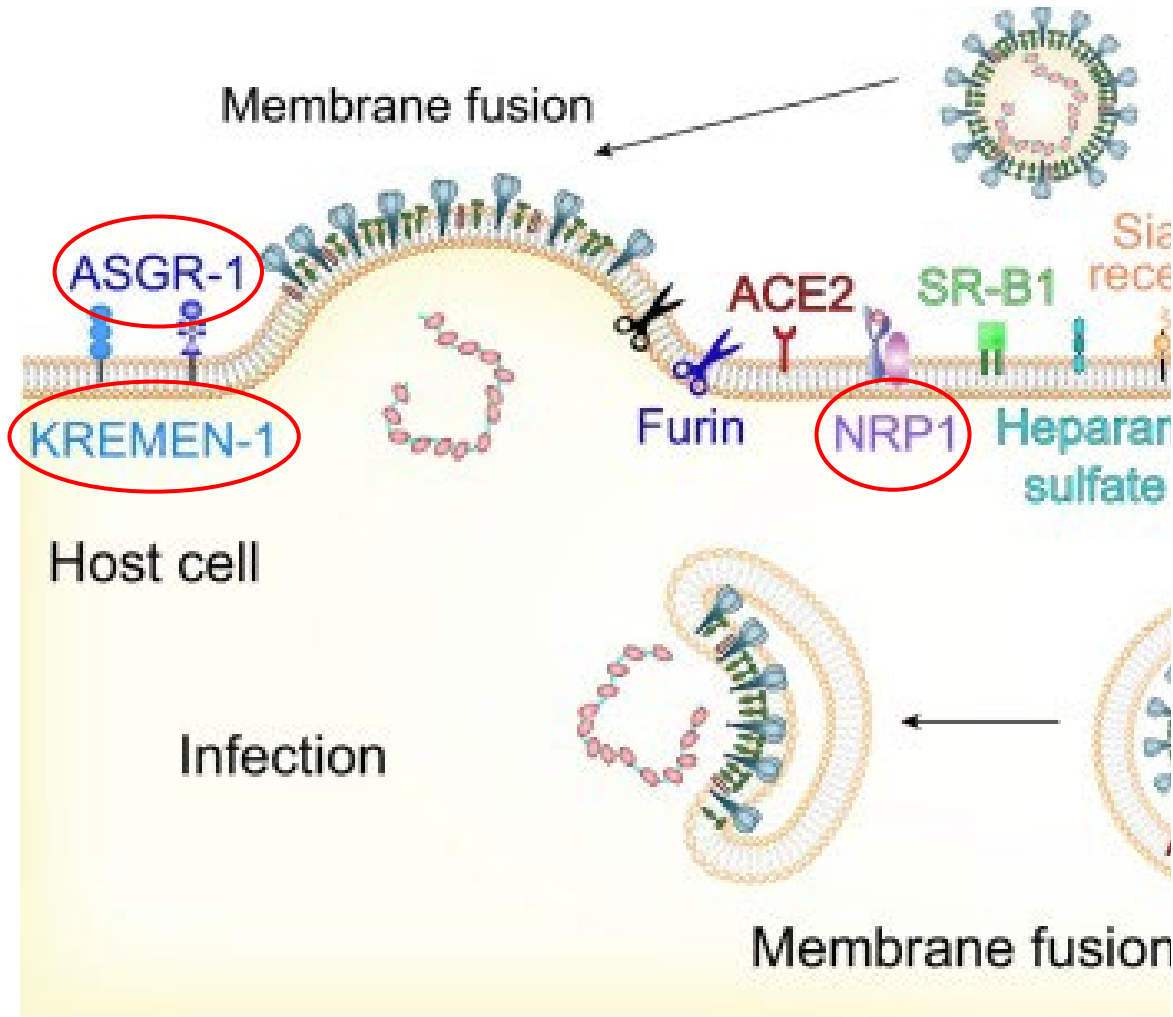


Table 1. Receptors, coreceptors, and cofactors involved in SARS-CoV-2 entry

Molecule	Category	Functional annotation
ACE2	Receptor	The major entry receptor
AXL	Receptor	A potential alternative receptor independent of ACE2
KREMEN1	Receptor	A potential alternative receptor independent of ACE2
ASGL1	Receptor	A potential alternative receptor independent of ACE2
CD147	Receptor	A potential alternative receptor independent of ACE2
Heparan sulfate	Coreceptor	Auxiliary attachment receptor, dependent on ACE2
Salic acid	Coreceptor	Auxiliary attachment receptor, dependent on ACE2
Lectin receptors	Coreceptor	Auxiliary attachment receptor, dependent on ACE2
Neuropilin 1	Coreceptor	Auxiliary attachment receptor, dependent on ACE2
CD4	Coreceptor	Potential auxiliary attachment receptor, dependent on ACE2
SR-B1/cholesterol	Cofactor	The S protein binds to cholesterol, and SR-B1 increases virion endocytosis by promoting cholesterol uptake
Furin	Cofactor	Proteolysis of S protein at the S1/S2 site
PC-1	Cofactor	Proteolysis of S protein at the S1/S2 site
Trypsin	Cofactor	Proteolysis of S protein at the S1/S2 site
Matriptase	Cofactor	Proteolysis of S protein at the S1/S2 site
Cathepsins	Cofactor	Proteolysis of S protein at the S1/S2 and S2' sites
TMPRSS2	Cofactor	Proteolysis of S protein at the S2' site

**alternative/auxiliary  
(co)receptors**

NRP1: neuropilin 1

KREMEN-1: Kringle Containing Transmembrane Protein 1

ASGR-1: Asialoglycoprotein receptor 1

Peng et al.

Trends Biochem Sci. 2021



May 5, 2020



## Review Article

### SARS-COV-2/COVID-19: EVOLVING REALITY, GLOBAL RESPONSE, KNOWLEDGE GAPS, AND OPPORTUNITIES

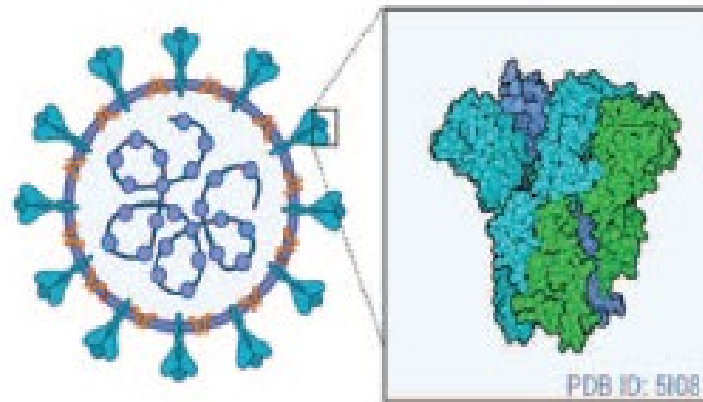
**Marcin F. Osuchowski,<sup>\*</sup> Federico Aletti,<sup>†</sup> Jean-Marc Cavillon,<sup>‡</sup>  
Stefanie B. Flohé,<sup>§</sup> Evangelos J. Giamarellos-Bourboulis,<sup>||</sup> Markus Huber-Lang,<sup>¶</sup>  
Borna Relja,<sup>#</sup> Tomasz Skirecki,<sup>\*\*</sup> Andrea Szabó,<sup>††</sup> and Marc Maegele<sup>‡‡§§</sup>**

*<sup>\*</sup>Ludwig Boltzmann Institute for Experimental and Clinical Traumatology in the AUVA Trauma Research Center, Vienna, Austria; <sup>†</sup>Department of Bioengineering, University of California San Diego, La Jolla, California; <sup>‡</sup>National Research Agency, Paris, France; <sup>§</sup>Department of Trauma, Hand, and Reconstructive Surgery, University Hospital Essen, University Duisburg-Essen, Essen, Germany; <sup>||</sup>4th Department of Internal Medicine, National and Kapodistrian University of Athens, Medical School, Athens, Greece; <sup>¶</sup>Institute of Clinical and Experimental Trauma-Immunology, University Hospital Ulm, Ulm University, Ulm, Germany; <sup>#</sup>Experimental Radiology, Department of Radiology and Nuclear Medicine, Otto von Guericke University Magdeburg, Magdeburg, Germany; <sup>\*\*</sup>Laboratory of Flow Cytometry, Centre of Postgraduate Medical Education, Warsaw, Poland; <sup>††</sup>Institute of Surgical Research, University of Szeged, Szeged, Hungary; <sup>‡‡</sup>Department of Trauma and Orthopaedic Surgery, Cologne-Merheim Medical Center (CMMC), University of Witten/Herdecke, Cologne-Merheim Campus, Cologne, Germany; and <sup>§§</sup>Institute for Research in Operative Medicine (IFOM), University of Witten/Herdecke, Cologne-Merheim Campus, Cologne, Germany*

# Defining “Goodness and Badness”

(on May 5, 2020)

- IFN I & III induction
- Coordinated monocyte response
- Induction of T<sub>fh</sub>
- Activation of ASCs
- Seroconversion



- Inflammasome activation
- Endothelial & epithelial injury
- Coagulopathy
- CTL overactivation
- Inflammatory monocyte
- Activation of BM monocytes
- Failure to generate protective Abs

Beneficial response

Pathological response

T<sub>fh</sub>: follicular helper T cells, **ASC**: antibody secreting cells, **CTL**: cytotoxic T-cells, **BM**: bone marrow

May 6, 2021

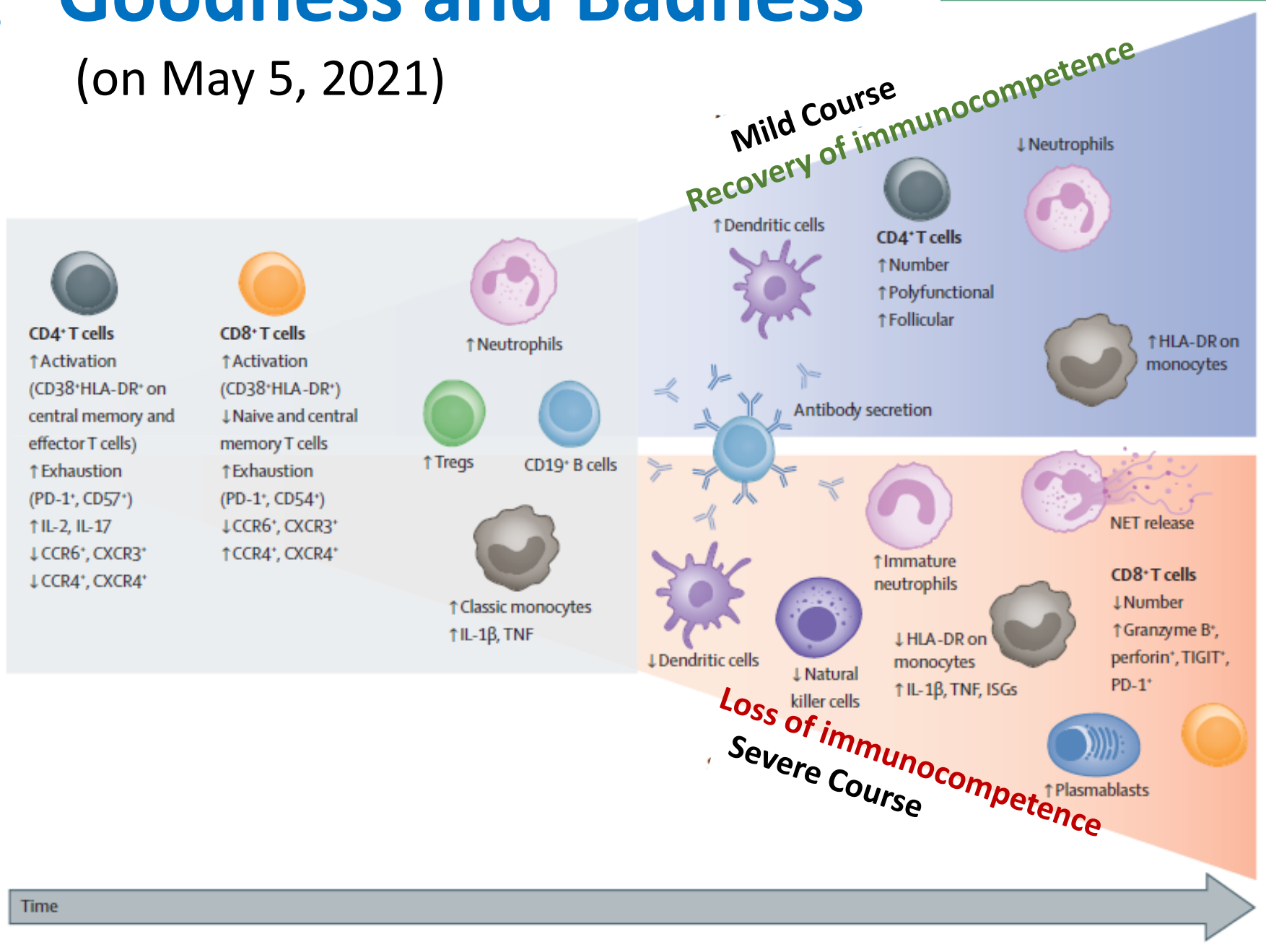
# COVID-19: Pathophysiology of Acute Disease 1

## The COVID-19 puzzle: deciphering pathophysiology and phenotypes of a new disease entity

*Marcin F Osuchowski\*, Martin S Winkler\*, Tomasz Skirecki, Sara Cajander, Manu Shankar-Hari, Gunnar Lachmann, Guillaume Monneret, Fabienne Venet, Michael Bauer, Frank M Brunkhorst, Sebastian Weis, Alberto Garcia-Salido, Matthijs Kox, Jean-Marc Cavaillon, Florian Uhle, Markus A Weigand, Stefanie B Flohé, W Joost Wiersinga, Raquel Almansa, Amanda de la Fuente, Ignacio Martin-Loeches, Christian Meisel, Thibaud Spinetti, Joerg C Schefold, Catia Cilloniz, Antoni Torres, Evangelos J Giamarellos-Bourboulis, Ricard Ferrer, Massimo Girardis, Andrea Cossarizza, Mihai G Netea, Tom van der Poll, Jesús F Bermejo-Martín, Ignacio Rubio*

# Defining “Goodness and Badness”

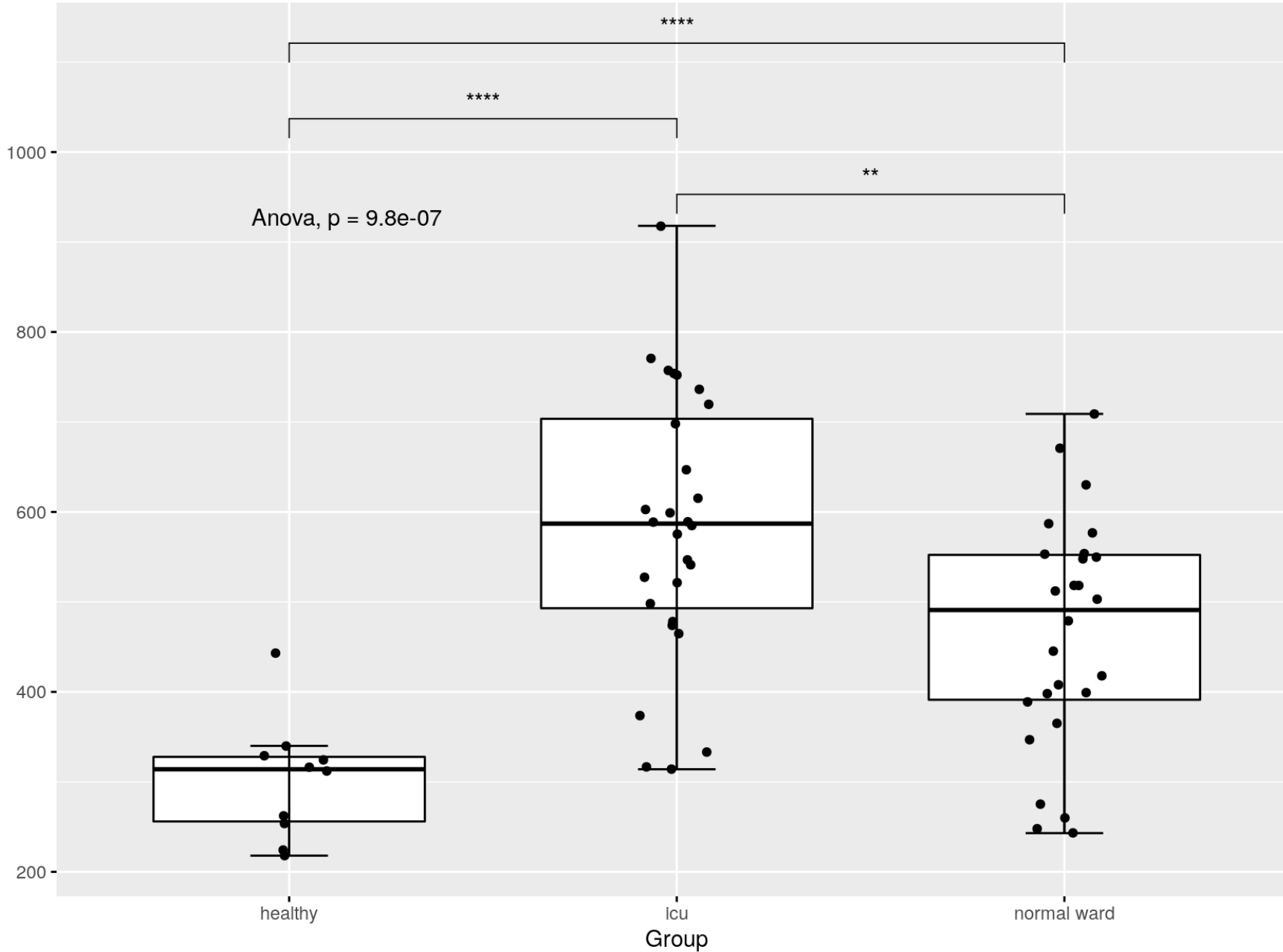
(on May 5, 2021)





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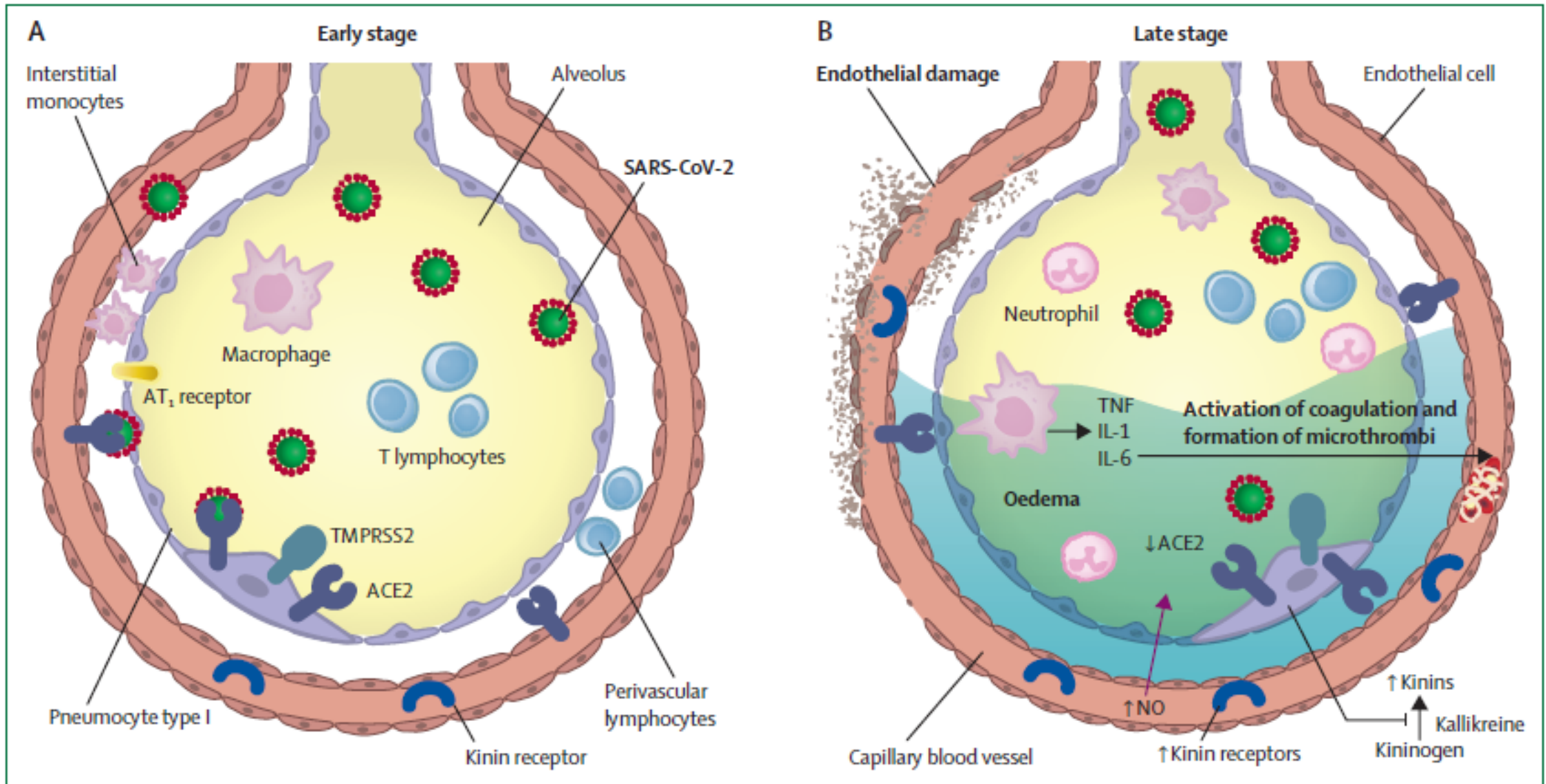
AUC: 0.7198



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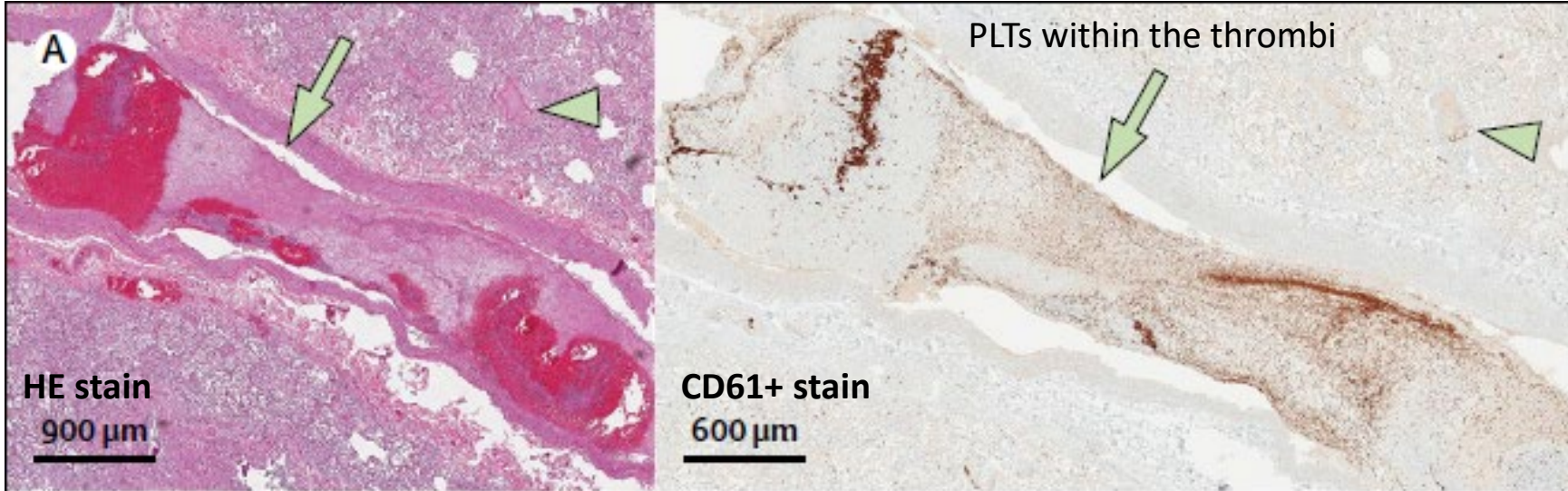
# Alveolar epithelial and endothelial damage, and coagulopathy in COVID-19



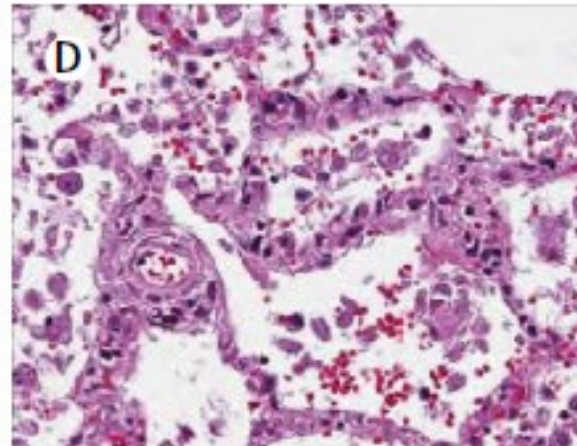
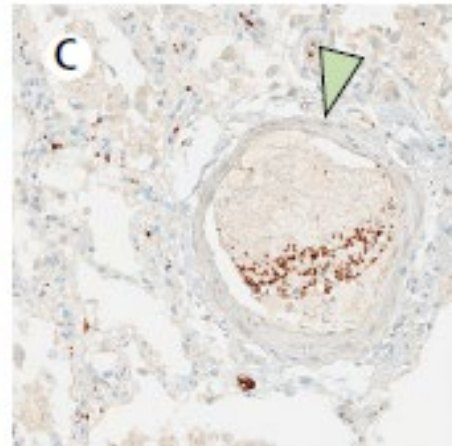
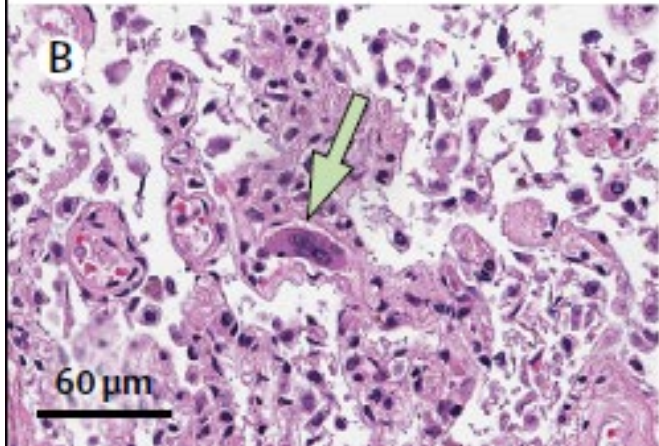
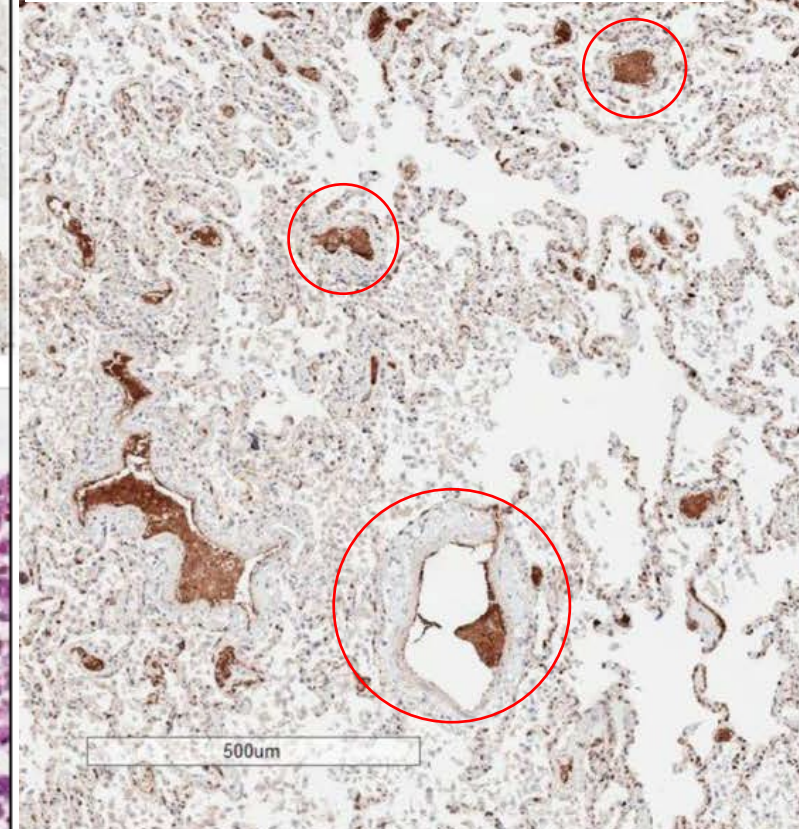


# Diffuse Alveolar Damage, Microangiopathy, Alveolar/Vascular Thrombosis

thrombus in a small pulmonary artery



vWF stain:  
small arteries/veins with accumulated  
PLTs & megakaryocytes



megakaryocytes in alveolar capillaries

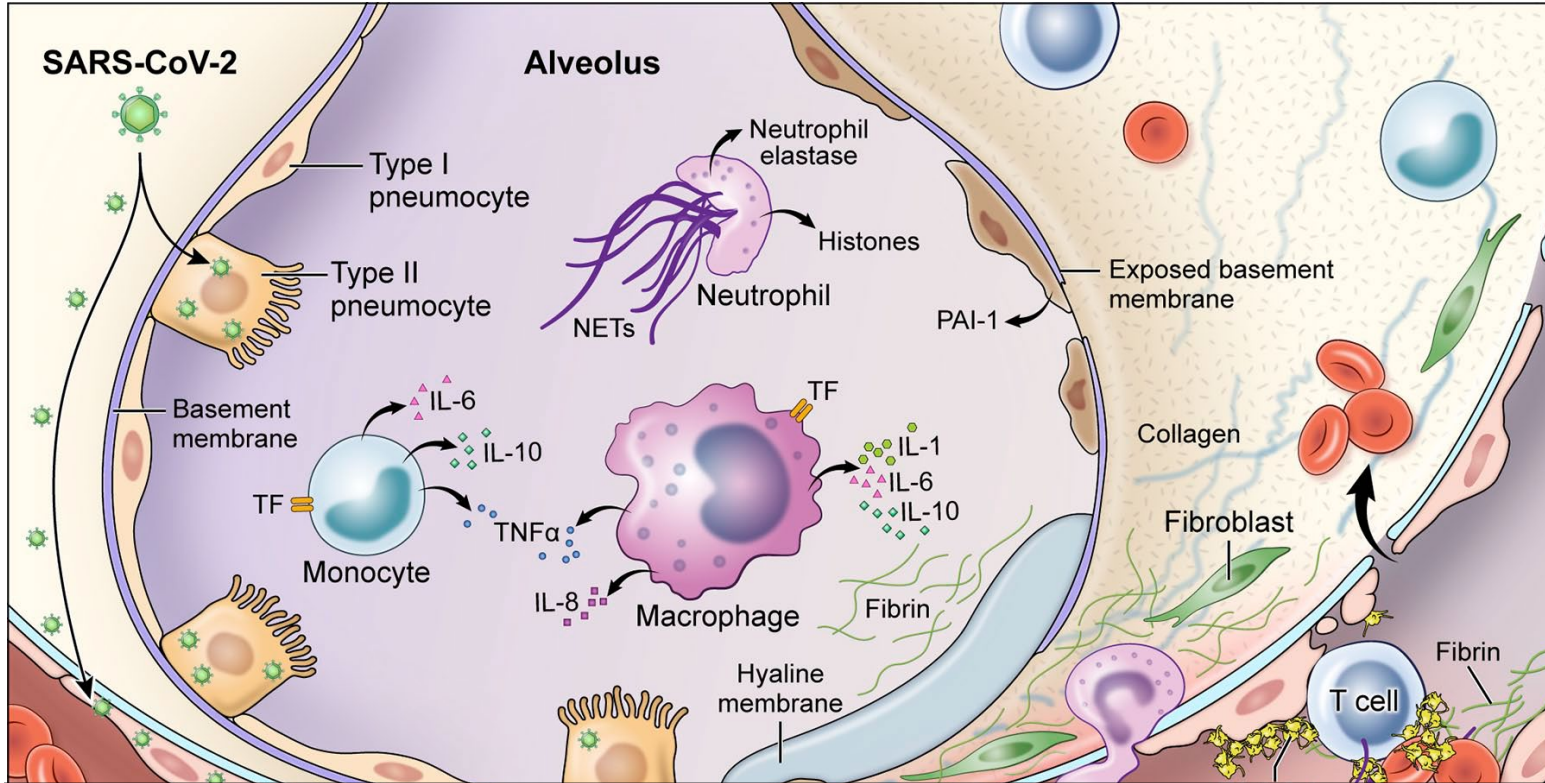
fibrin/PLT thrombus

perivascular LYM aggregates



# Coagulopathy in the Alveolus & Capillary

Immune activation and mechanisms of coagulopathy



## NOT a Classical DIC!

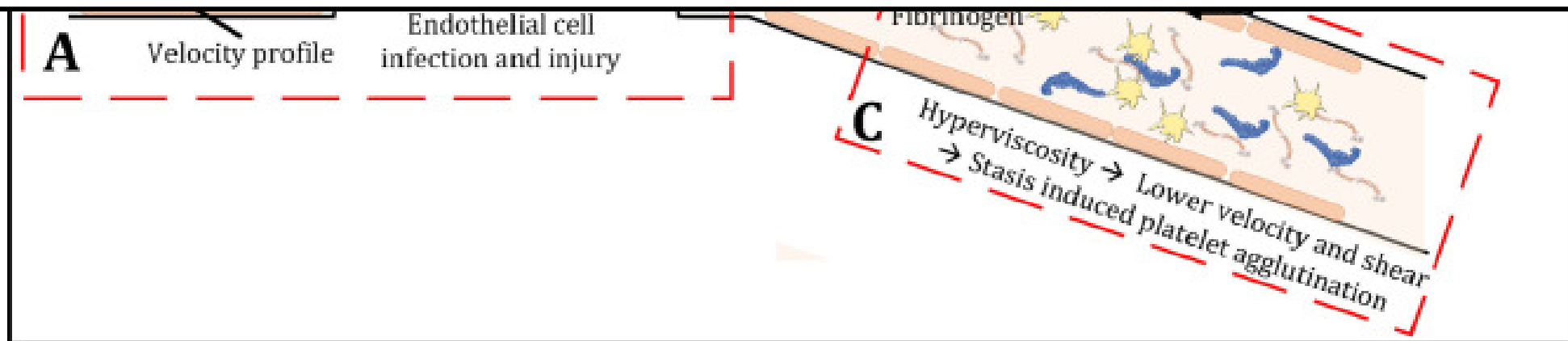
- mildly/not prolonged PT/aPTT
- Mild thrombocytopenia
- High/very high fibrinogen
  - High D-dimers



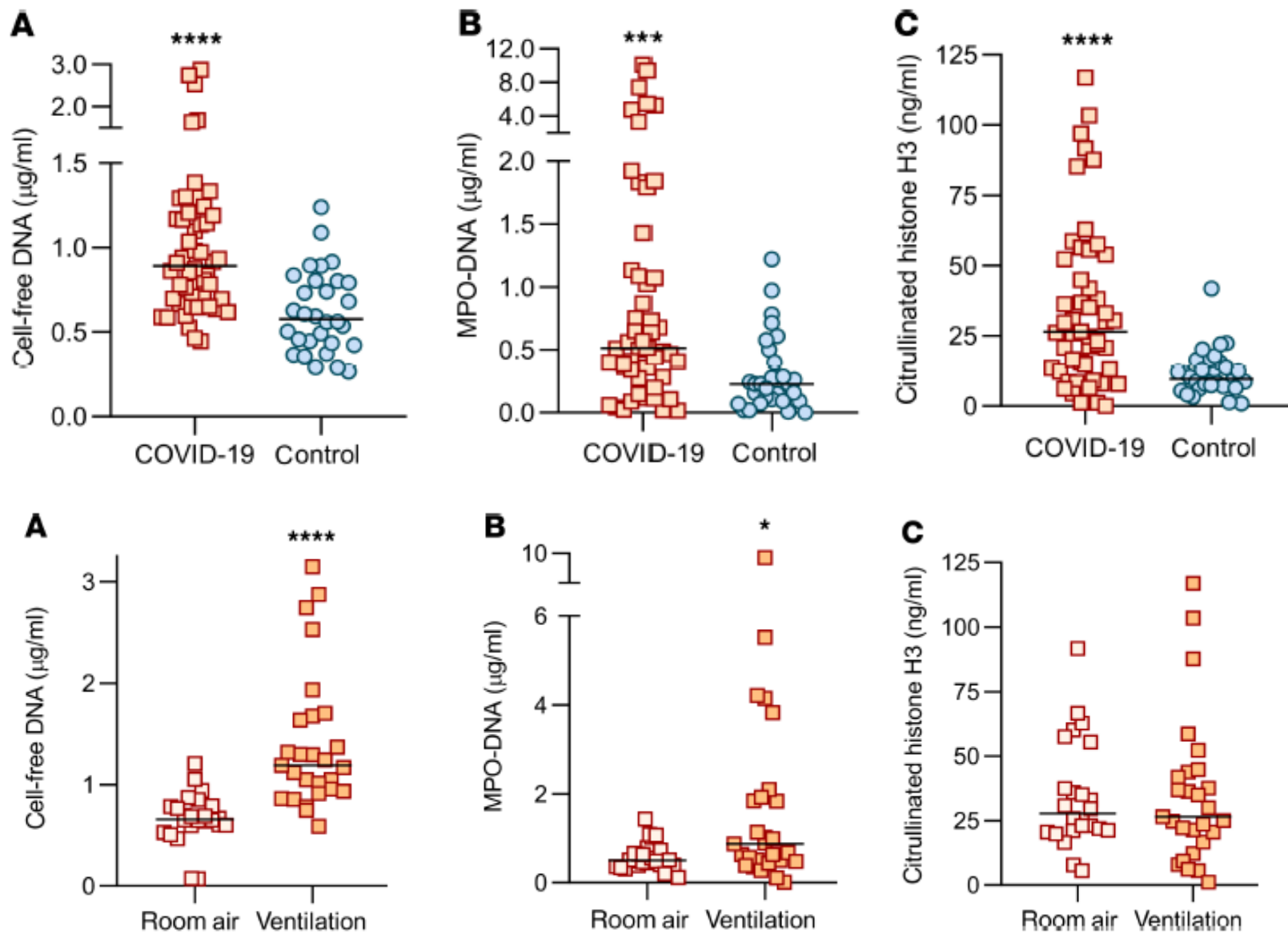
# COVID-19 and thrombosis: The role of hemodynamics



**SARS-CoV-2-induced endotheliitis is a common factor in the respiratory and non-respiratory manifestations of COVID-19**

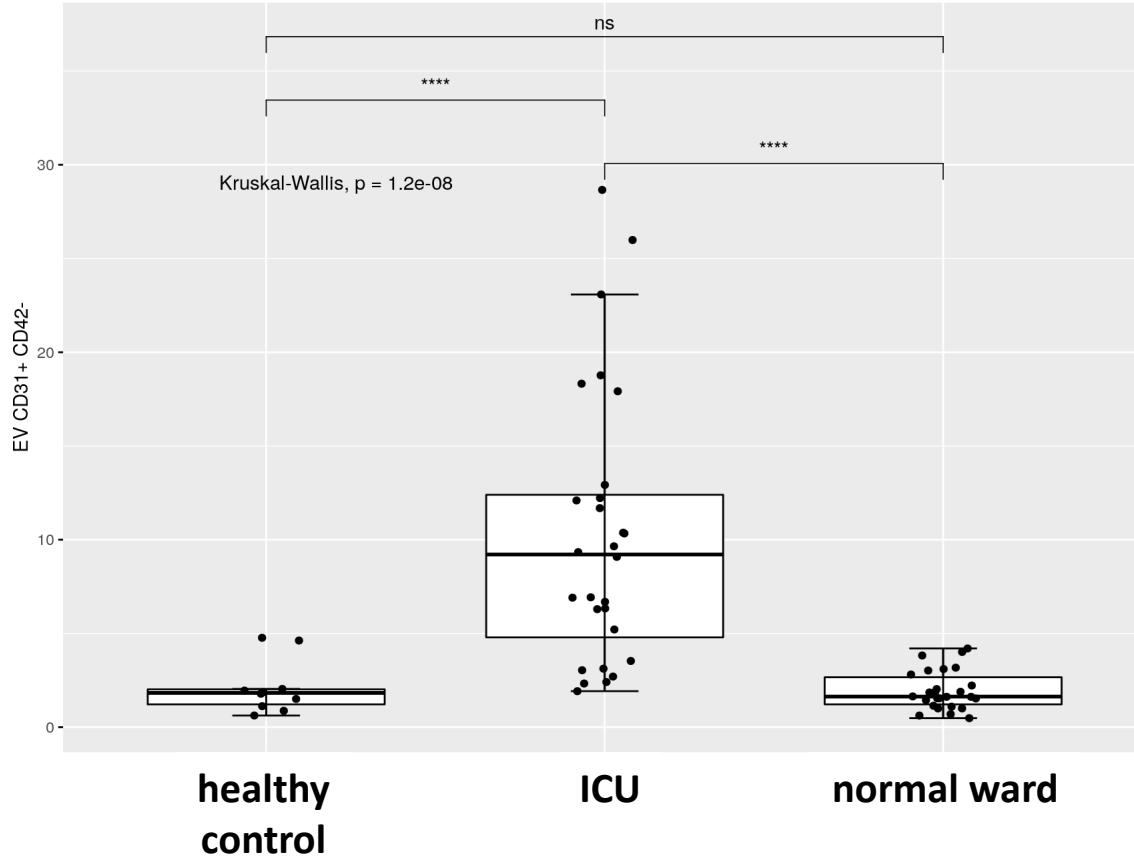


# Neutrophil extracellular traps in COVID-19

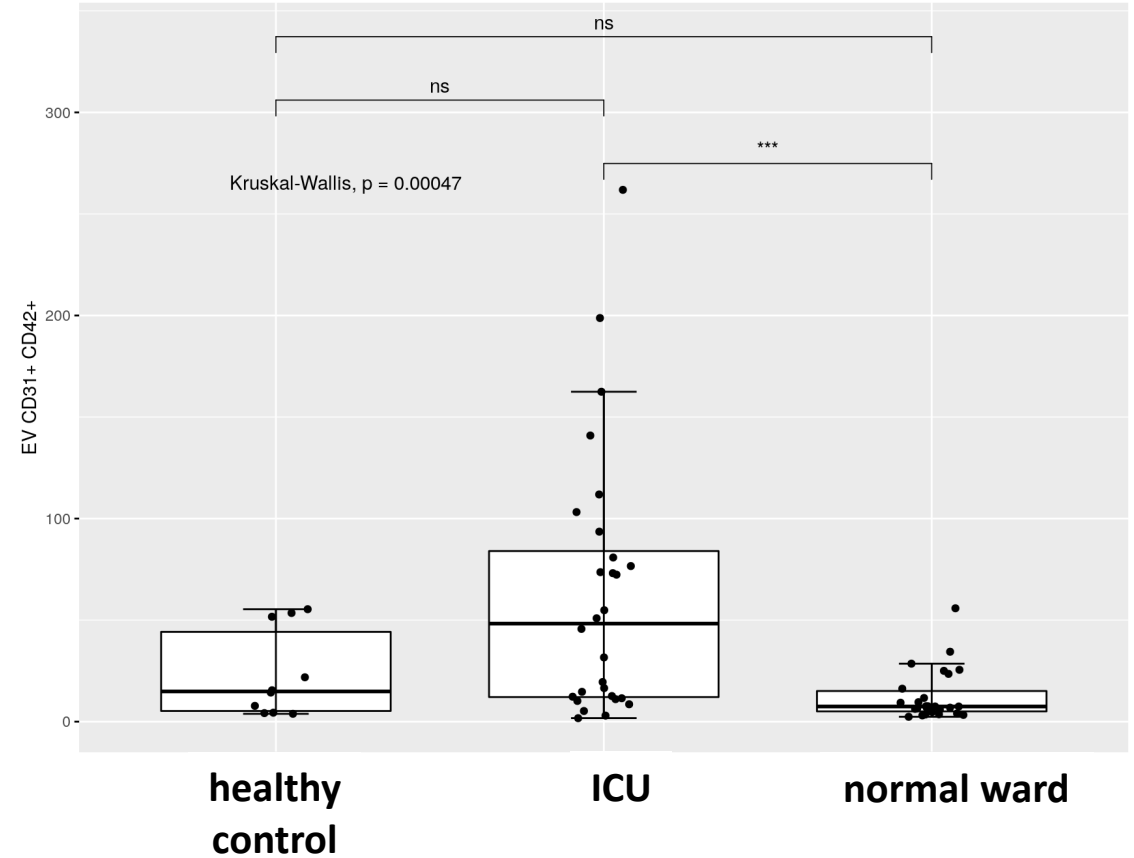


# High Release of Endoth./PLT-derived Extracellular Vesicles in Severe C-19

EV CD31+ endothelium  
AUC: 0.9423



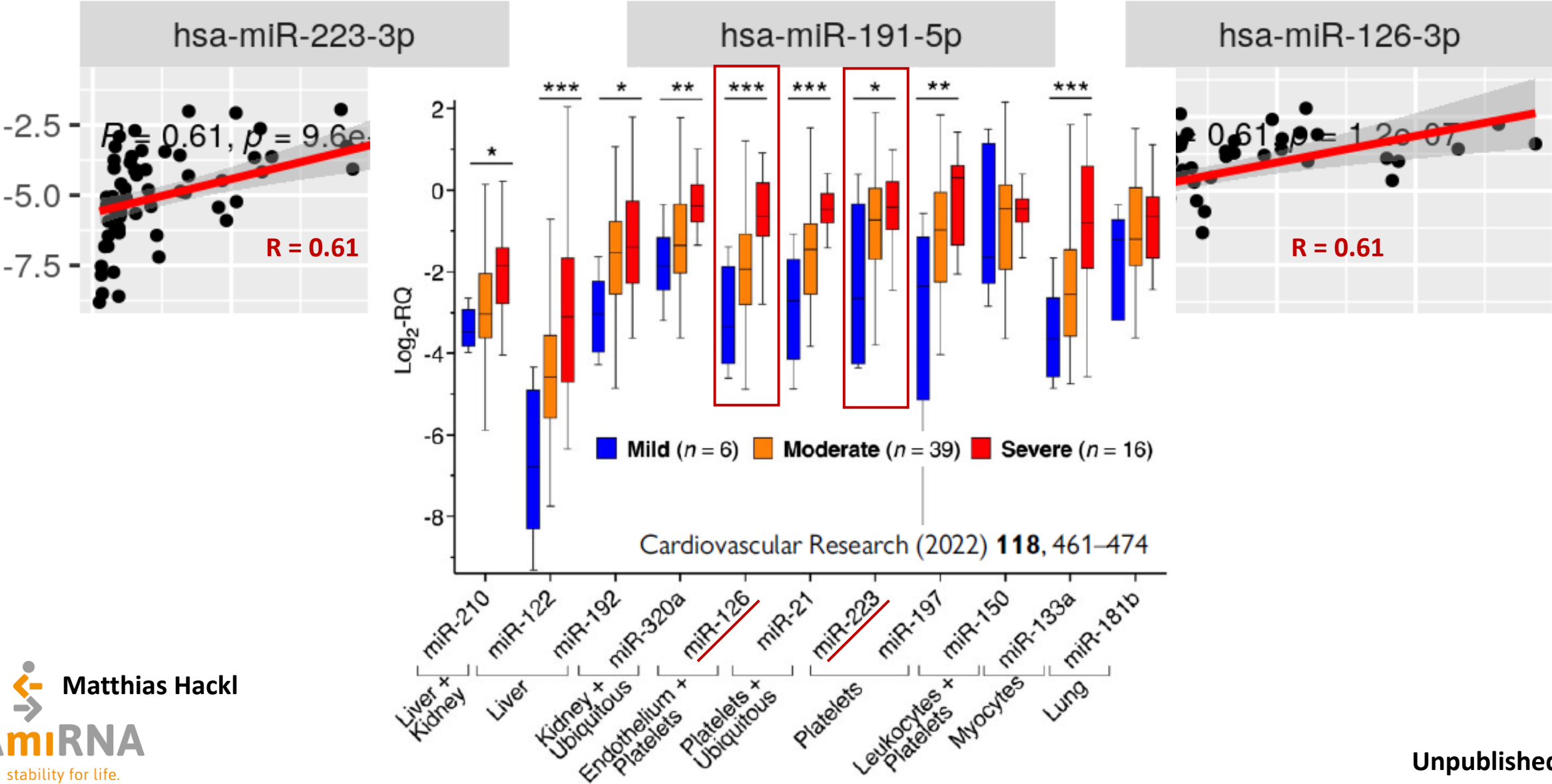
EV CD31+ CD42+ platelets  
AUC: 0.8049



Eva Schaden  
Johannes Gratz

Marion Wiegele  
Pierre Raeven

# microRNA signal well-correlated with the EV load in COVID-19 patients





# A COVID-19 Cytokine Storm??



JCI INSIGHT

2020;5(17):e140329.

## Severe immunosuppression and not a cytokine storm characterizes COVID-19 infections

Kenneth E. Remy,<sup>1,2,3</sup> Monty Mazer,<sup>1,3</sup> David A. Striker,<sup>4</sup> Ali H. Ellebedy,<sup>5</sup> Andrew H. Walton, Jacqueline Unsinger,<sup>3</sup> Teresa M. Blood,<sup>3</sup> Philip A. Mudd,<sup>6</sup> Daehan J. Yi,<sup>1</sup> Daniel A. Mannion,<sup>1</sup> Dale F. Osborne,<sup>3</sup> R. Scott Martin,<sup>4</sup> Nitin J. Anand,<sup>4</sup> James P. Bosanquet,<sup>4</sup> Jane Blood,<sup>3</sup> Anne M. Drewry,<sup>3</sup> Charles C. Caldwell,<sup>8</sup> Isaiah R. Turnbull,<sup>9</sup> Scott C. Brakenridge,<sup>10</sup> Lyle L. Moldwaver,<sup>10</sup> and Richard S. Hotchkiss<sup>2,3,9</sup>

Lancet Respir Med 2020;

8: 1233–44

## Cytokine elevation in severe and critical COVID-19: a rapid systematic review, meta-analysis, and comparison with other inflammatory syndromes

Daniel E Leisman\*, Lukas Ronner\*, Rachel Pinotti, Matthew D Taylor, Pratik Sinha, Carolyn S Calfee, Alexandre V Hirayama, Fiore Mastroianni, Cameron J Turtle, Michael O Harhay, Matthieu Legrand, Clifford S Deutschman

JOURNAL OF  
MEDICAL VIROLOGY WILEY

J Med Virol. 2020;1–2.

## COVID-19: What type of cytokine storm are we dealing with?

JAMA Internal Medicine September 2020 Volume 180, Number 9

EDITORIAL

### Is a “Cytokine Storm” Relevant to COVID-19?

Pratik Sinha, MB, ChB, PhD; Michael A. Matthay, MD; Carolyn S. Calfee, MD, MAS

SHOCK, Vol. 56, No. 5, pp. 667–672, 2021

Review Article

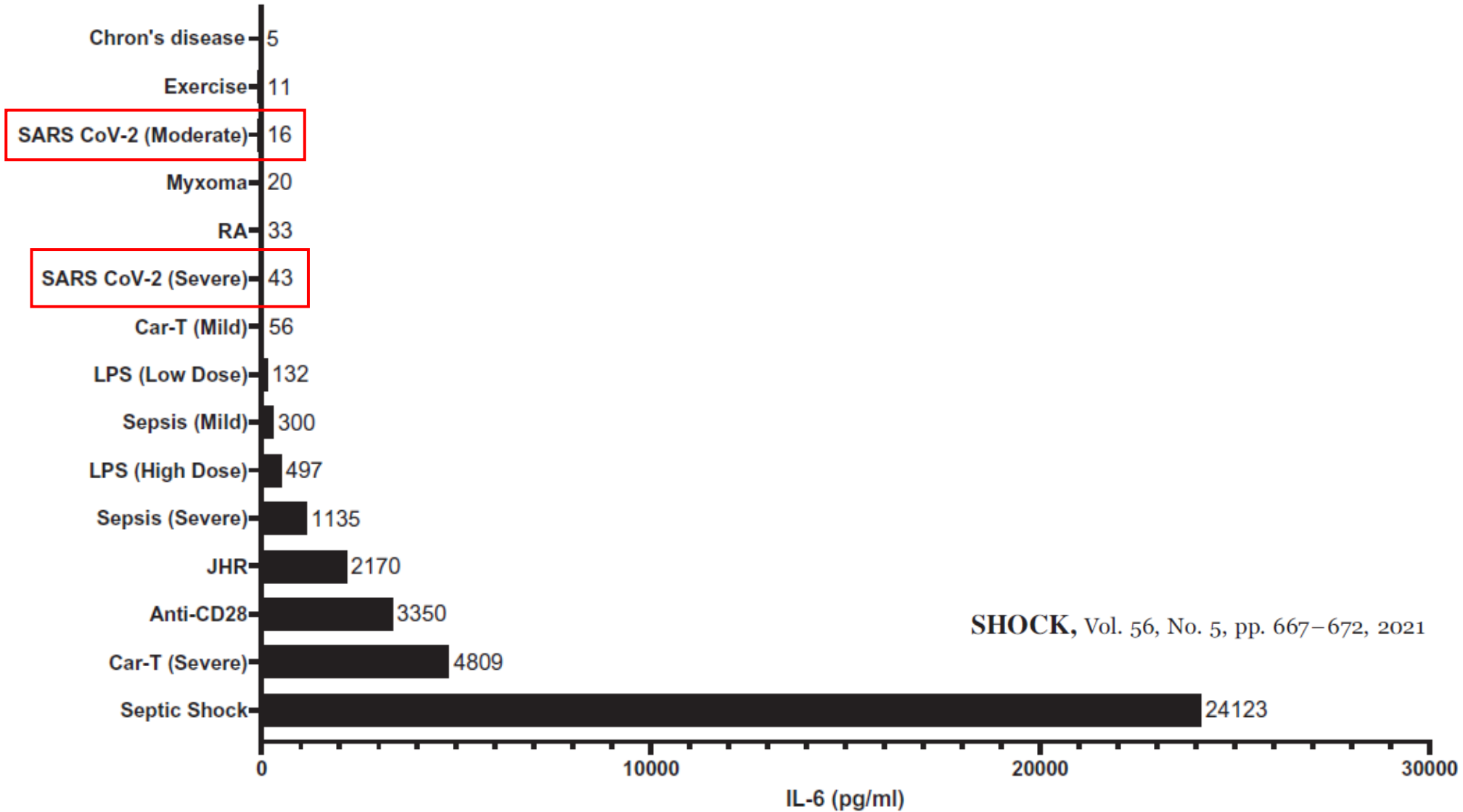
## CYTOKINE DRIZZLE—THE RATIONALE FOR ABANDONING “CYTOKINE STORM”

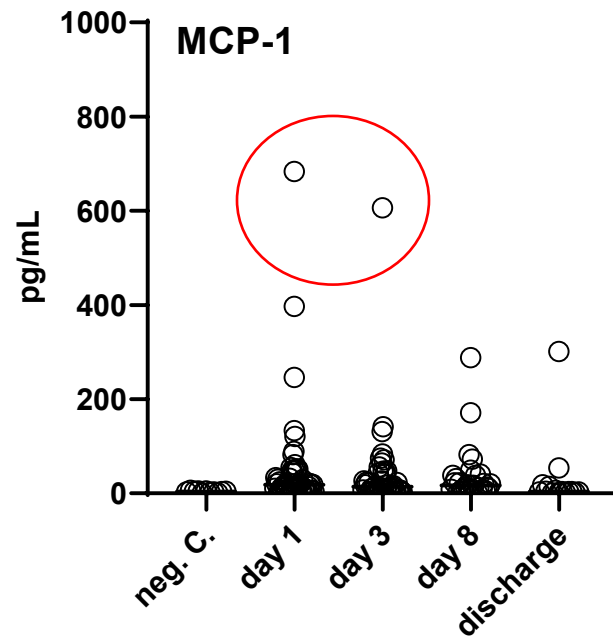
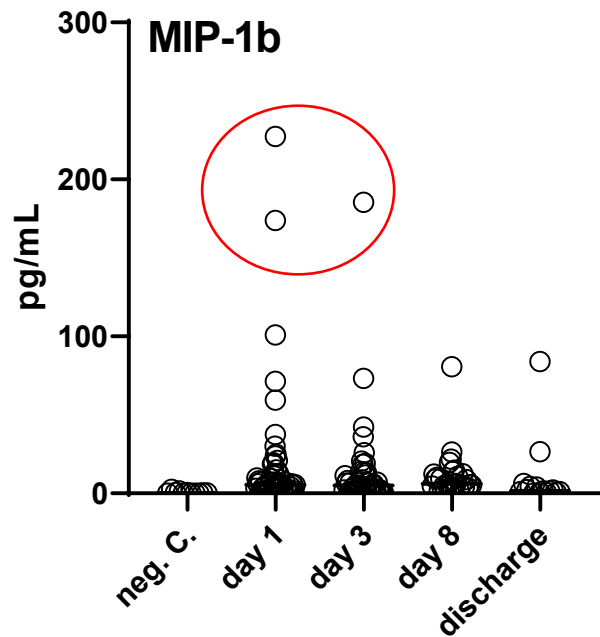
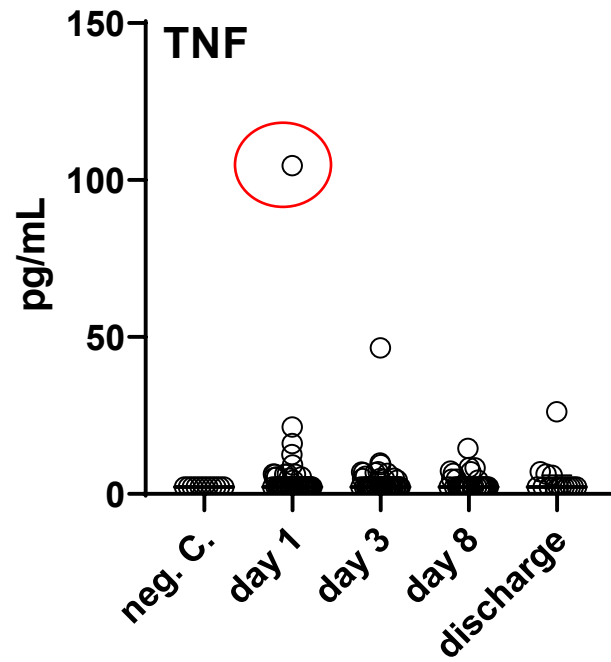
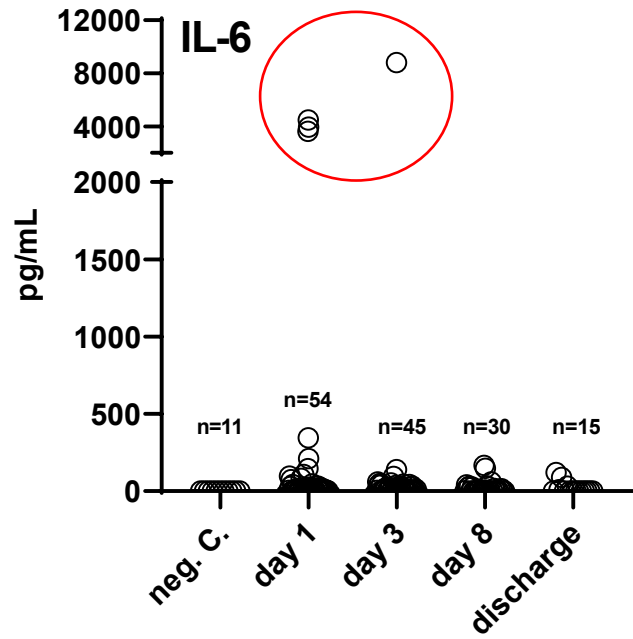
Allan E. Stolarski,<sup>\*†</sup> Jiyoun Kim,<sup>†</sup> Qiuyang Zhang,<sup>†</sup> and Daniel G. Remick<sup>†</sup>

<sup>\*</sup>Department of Surgery, Boston Medical Center, Boston University, Boston, Massachusetts; and

<sup>†</sup>Department of Pathology and Laboratory Medicine, Boston University, Boston, Massachusetts

# Circulating IL-6 Across Various Severe Conditions





# Cytokine Storm??

Damn! I can't reflexively believe in everything I read on PubMed...

There is NO cytokine storm in COVID-19!!! How many times do I have to repeat it?!

Need a journal club on this and FAST!

COVID-19?!  
What's that?!?

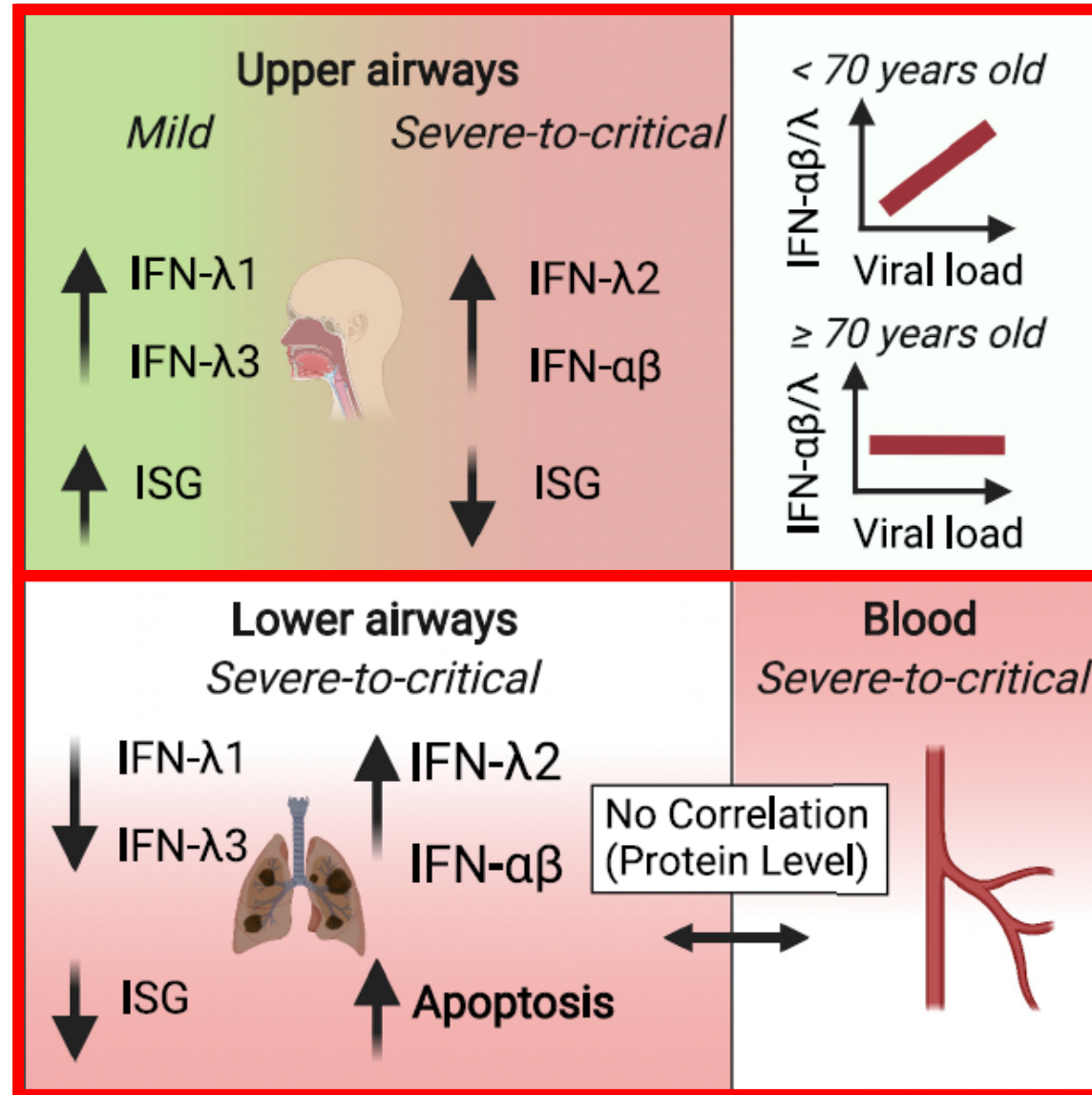
Sooo glad I read the recent Shock review on the topic!

**NOT Really!!**





# The interferon landscape along the respiratory tract impacts the severity of COVID-19

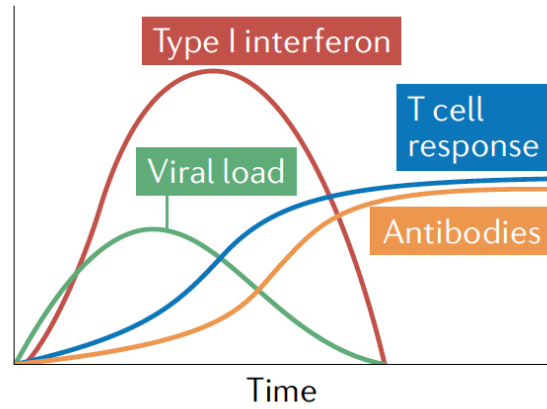
Alpha/beta ( $\alpha/\beta$ ): IFN Type I  
 Lambda ( $\lambda$ ): IFN Type III



## The first 12 months of COVID-19: a timeline of immunological insights

Thiago Carvalho, Florian Krammer  and Akiko Iwasaki 

**a** Early robust type I interferon response

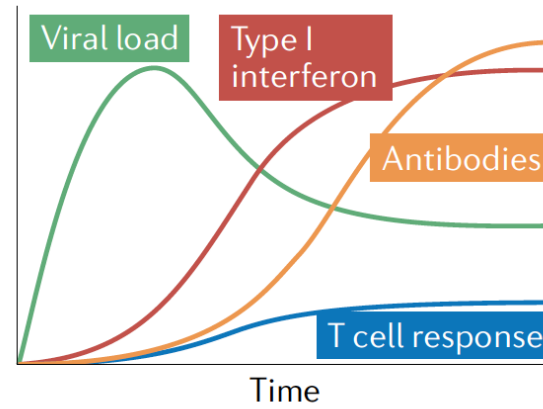


- Viral clearance
- Normal-level T cell and B cell responses

Mild disease

- Young adults
- Low levels of viral exposure

**b** Delayed type I interferon response

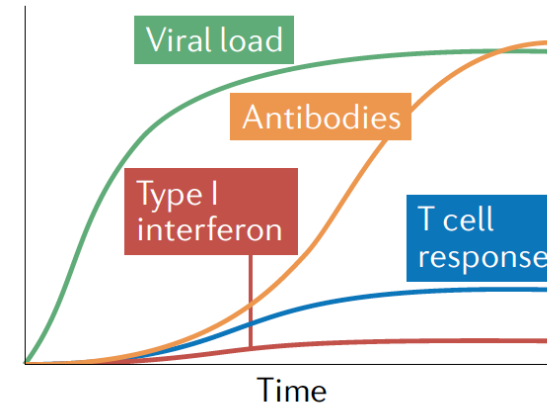


- Partial viral clearance
- T cell lymphopenia; robust B cell response

Severe disease

- Older adults
- Higher levels of viral exposure

**c** Type I interferon deficiency

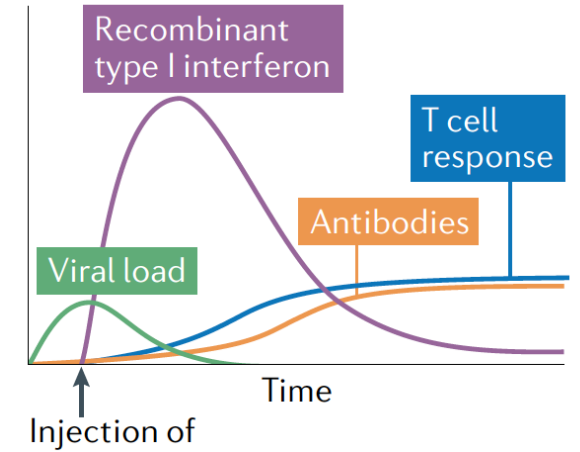


- Uncontrolled viral replication
- T cell lymphopenia; compensatory B cell response

Severe disease

- Genetic mutations in type I interferon pathways
- Neutralizing antibodies to type I interferons

**d** Recombinant type I interferon therapy



- Rapid viral clearance
- Reduced T cell and B cell responses

Milder disease

- Early treatment with recombinant type I interferon

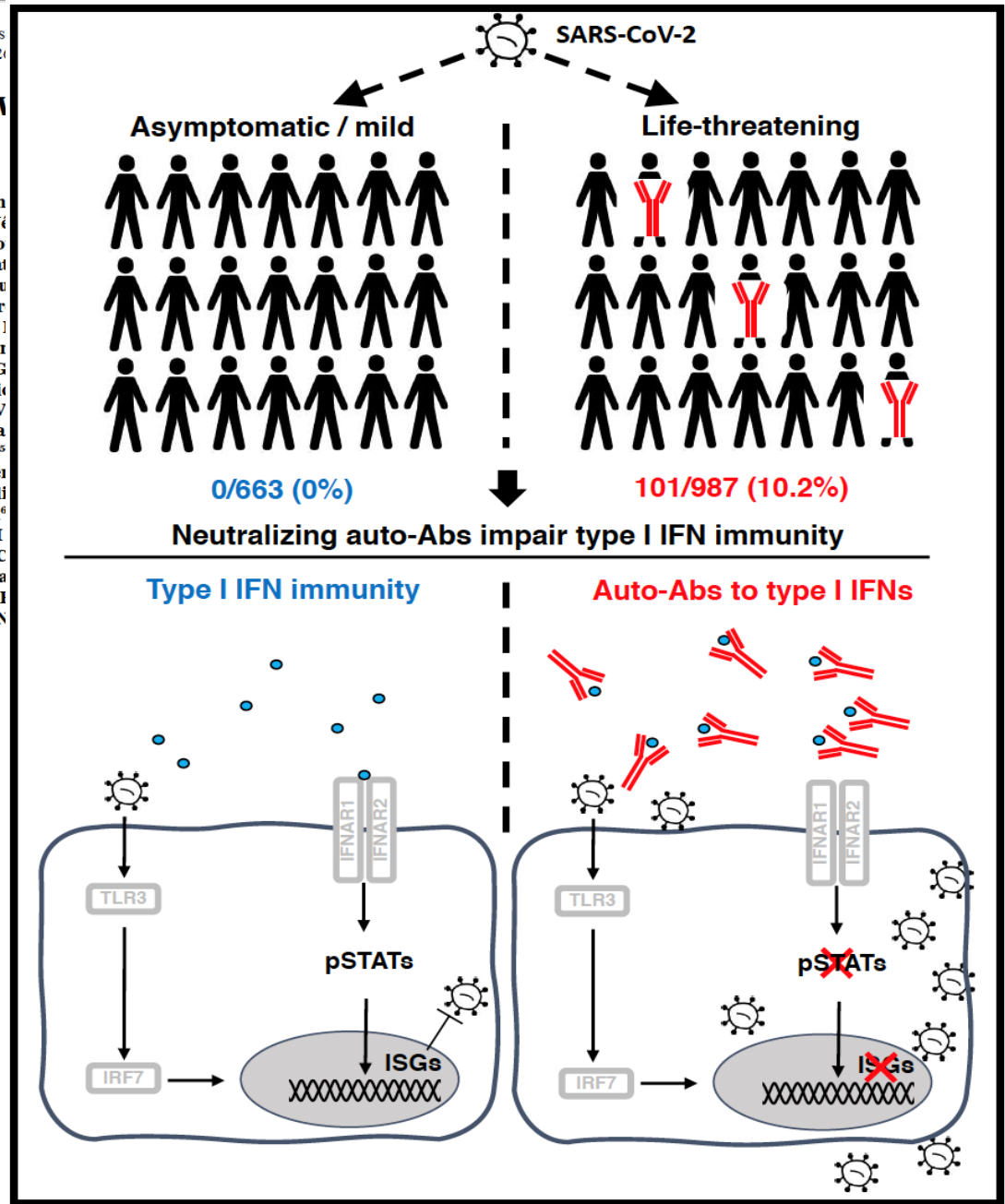
**GENETIC PREDISPOSITION**

**Auto-antibodies against type I IFNs in patients with threatening COVID-19**

Paul Bastard<sup>1,2,3,4\*</sup>, Lindsey B. Rosen<sup>4†</sup>, Qian Zhang<sup>3,4</sup>, Eleftherios Michailidis<sup>5,4</sup>, Hans-Hein Zhang<sup>4†</sup>, Karim Dorgham<sup>6†</sup>, Quentin Philippot<sup>1,2,4</sup>, Jérémie Rosain<sup>1,2,4</sup>, Vivien Béziat<sup>1,2,3,4</sup>, Jé Elana Shaw<sup>4</sup>, Liis Haljasmägi<sup>7</sup>, Pärt Peterson<sup>7</sup>, Lazaro Lorenzo<sup>1,2</sup>, Lucy Bizien<sup>1,2</sup>, Sophie Tro Kerry Dobbs<sup>4</sup>, Adriana Almeida de Jesus<sup>4</sup>, Alexandre Belot<sup>10,11,12</sup>, Anne Kallaste<sup>1,3</sup>, Emilie Cat Tandjaoui-Lambiotte<sup>15</sup>, Jeremie Le Pen<sup>5</sup>, Gaspard Kerner<sup>1,2</sup>, Benedetta Bigio<sup>3</sup>, Yoann Seeleu Alexandre Bolze<sup>16</sup>, Andrés N. Spaan<sup>3,17</sup>, Ottavia M. Delmonte<sup>4</sup>, Michael S. Abers<sup>4</sup>, Alessandr Casari<sup>18</sup>, Vito Lampasona<sup>18</sup>, Lorenzo Piemonti<sup>18</sup>, Fabio Ciceri<sup>18</sup>, Kaya Bilguvar<sup>19</sup>, Richard P. J Vasse<sup>22</sup>, David M. Smadja<sup>23</sup>, Mélanie Migaud<sup>1,2</sup>, Jérôme Hadjadj<sup>24</sup>, Benjamin Terrier<sup>25</sup>, Dari Quintana-Murci<sup>27,28</sup>, Diederik van de Beek<sup>29</sup>, Lucie Roussel<sup>30,31</sup>, Donald C. Vinh<sup>30,31</sup>, Stuart G Filomeen Haerynck<sup>34</sup>, David Dalmau<sup>35</sup>, Javier Martínez-Picado<sup>36,37,38</sup>, Petter Brodin<sup>39,40</sup>, Mi Nussenzweig<sup>41,42</sup>, Stéphanie Boisson-Dupuis<sup>1,2,3</sup>, Carlos Rodríguez-Gallego<sup>43,44</sup>, Guillaume V Mogensen<sup>46,47</sup>, Andrew J. Oler<sup>48</sup>, Jingwen Gu<sup>48</sup>, Peter D. Burbelo<sup>49</sup>, Jeffrey Cohen<sup>50</sup>, Andrea Rachele Bettini<sup>51</sup>, Mariella D'Angio<sup>51</sup>, Paolo Bonfanti<sup>52</sup>, Patrick Rossignol<sup>53</sup>, Julien Mayaux<sup>5</sup> Laucat<sup>24</sup>, Eystein S. Husebye<sup>55,56,57</sup>, Francesca Fusco<sup>58</sup>, Matilde Valeria Ursini<sup>58</sup>, Luisa Imber Sottini<sup>59</sup>, Simone Paghera<sup>59</sup>, Eugenia Quiros-Roldan<sup>60</sup>, Camillo Rossi<sup>61</sup>, Riccardo Castagnoli Montagna<sup>63,64</sup>, Amelia Licari<sup>62</sup>, Gian Luigi Marseglia<sup>62</sup>, Xavier Duval<sup>65,66,67,68,69</sup>, Jade Ghosn<sup>6</sup> NIAID-USUHS Immune Response to COVID Group§, COVID Clinicians§, COVID-STORM COVID Group§, French COVID Cohort Study Group§, The Milieu Intérieur Consortium§, C Amsterdam UMC Covid-19 Biobank§, COVID Human Genetic Effort§, John S. Tsang<sup>70,71</sup>, Ra Mansky<sup>4</sup>, Kai Kisand<sup>7</sup>, Michail S. Lionakis<sup>4</sup>, Anne Puel<sup>1,2,3</sup>, Shen-Ying Zhang<sup>1,2,3</sup>, Steven M. I Gorochov<sup>6,72¶</sup>, Emmanuelle Jouanguy<sup>1,2,3¶</sup>, Charles M. Rice<sup>5¶</sup>, Aurélie Cobat<sup>1,2,3¶</sup>, Luigi D. N Laurent Abel<sup>1,2,3¶</sup>, Helen C. Su<sup>4#</sup>, Jean-Laurent Casanova<sup>1,2,3,42,73#</sup>



**“At least 10% of critical COVID-19 is an autoimmune attack.”**  
Jean-Laurent Casanova, Rockefeller University



# COVID-19 Consequences Can Be Widespread/Long-term...

Body organs/parts	Damage/Consequences	Acts on/ Type of cell affected
<b>Gut Microbiota</b>	Gut dysbiosis	Decrease probiotic microorganisms
	Inflammation and diarrhoea	The small intestine, enterocytes, colon
<b>Kidney</b>	Acute Kidney Injury (AKI)	Podocytes and proximally straight tubular cells
	Rhabdomyolysis, hypoxemia, and coagulopathy	Glomerular cells, tubular epithelium, and podocytes
<b>Liver</b>	Irregular liver function	Cells of the bile duct
<b>Heart</b>	Microvascular disorders	Pericytes
	Myocarditis	Myocardium
	Acute Coronary Syndrome	Endothelial cells
<b>Lungs</b>	Alveolar damage, hypoxemia	Alveolar cell
	Acute Respiratory Distress Syndrome	Hyper fusion of alveolar cells
<b>Brain</b>	Cerebral haemorrhage	ACE-2 receptor affected due to amplified blood pressure
	Encephalitis	Neuronal destruction and nerve tissue lesions
	Puzzlement, loss of awareness, coma	Cerebral capillary endothelial cells



# Clinical characteristics with inflammation profiling of long COVID and association with 1-year recovery following hospitalisation in the UK: a prospective observational study

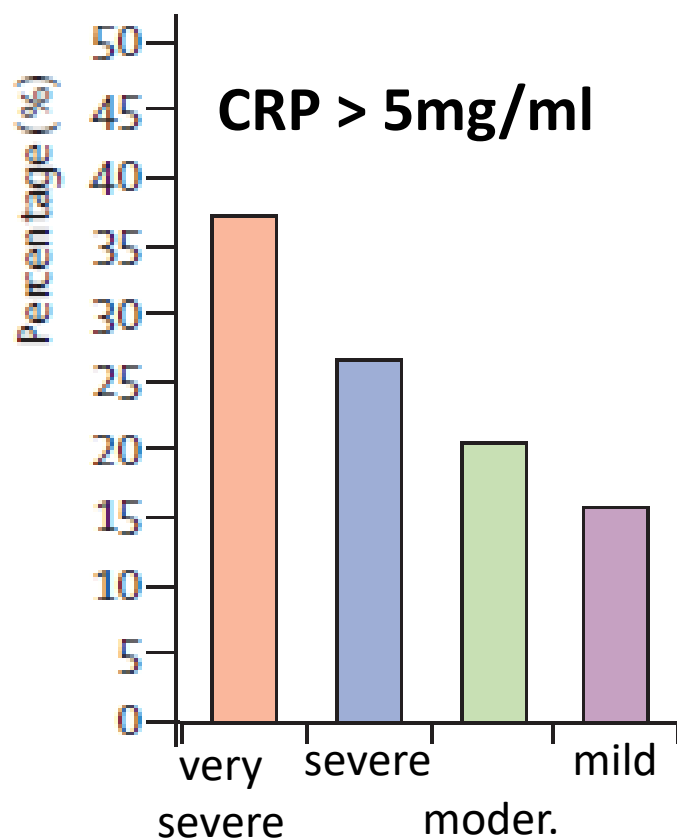
Lancet Respir Med 2022

Published Online

April 23, 2022

The PHOSP-COVID Collaborative Group\*

## Covid-19: Only a quarter of patients admitted to hospital feel fully recovered after a year, study finds



The ten most common persistent symptoms at 1 year after discharge were fatigue (463 [60.1%] of 770 patients), aching muscles (442 [54.6%] of 809), physically slowing down (429 [52.9%] of 811), poor sleep (402 [52.3%] of 769), breathlessness (395 [51.4%] of 769), joint pain or swelling (382 [47.6%] of 803), slowing down in thinking (377 [46.7%] of 808), pain (359 [46.6%] of 770), short-term memory loss (360 [44.6%] of 808), and limb weakness (341 [41.9%] of 813; appendix p 35). Overall,












## Panel 2: Priorities for future research

The proposed research aims will be achieved most effectively with a complementary combination of preclinical and clinical research.

- Establish the molecular basis for lower pathogenicity of SARS-CoV-2 compared with SARS-CoV
- Define the role of pre-existing and acquired T-cell immunity in COVID-19 development and progression
- Establish precise predictive thresholds for known biomarkers of COVID-19 severity, outcomes, and complications
- Develop novel prognostic biomarkers and risk predictors for COVID-19 pneumopathy, acute respiratory distress syndrome, and fibrosis
- Elucidate compartmentalisation profiles of soluble inflammatory mediators and cell subsets (ie, in individual organs and systems)
- Characterise immunological deficiencies secondary to ageing and comorbidities that impair efficient immunological responses against SARS-CoV-2
- Characterise short-term and long-term COVID-19 vasculopathies and their sequelae
- Develop a unified post-COVID-19 monitoring platform to characterise long-term outcomes and immune derangements after SARS-CoV-2 infection
- Conduct high-quality, prospective clinical studies to identify optimum anti-coagulative and immunomodulatory strategies for patients with SARS-CoV-2 infection

# A Wishful Thinking To-Do List...

-  some evidence emerging
-  some evidence emerging
-  robust evidence emerging; needs standardization
-  little evidence emerging
-  v. difficult; mostly evidence from blood/lung
-  some evidence emerging; need of Big Data
-  Robust evid. for short-term; poor for long-term
-  Difficult to organize; some local initiatives
-  mixed quality evidence: both robust & poor RCTs



**Thanks**

**Thanks**

**Thanks**

**Thanks**







20th  
Congress



September 21-23, 2023

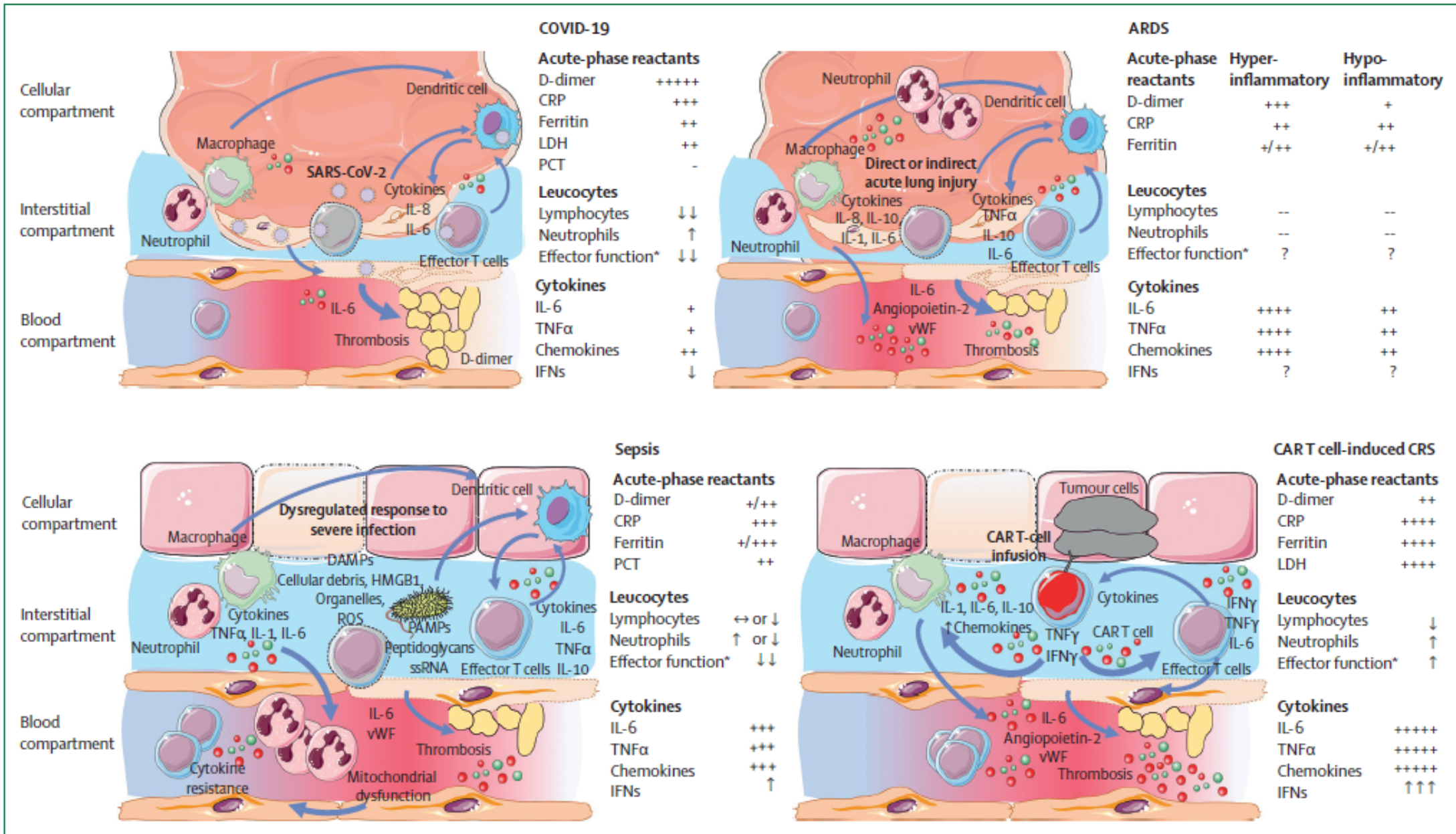


University of Vienna



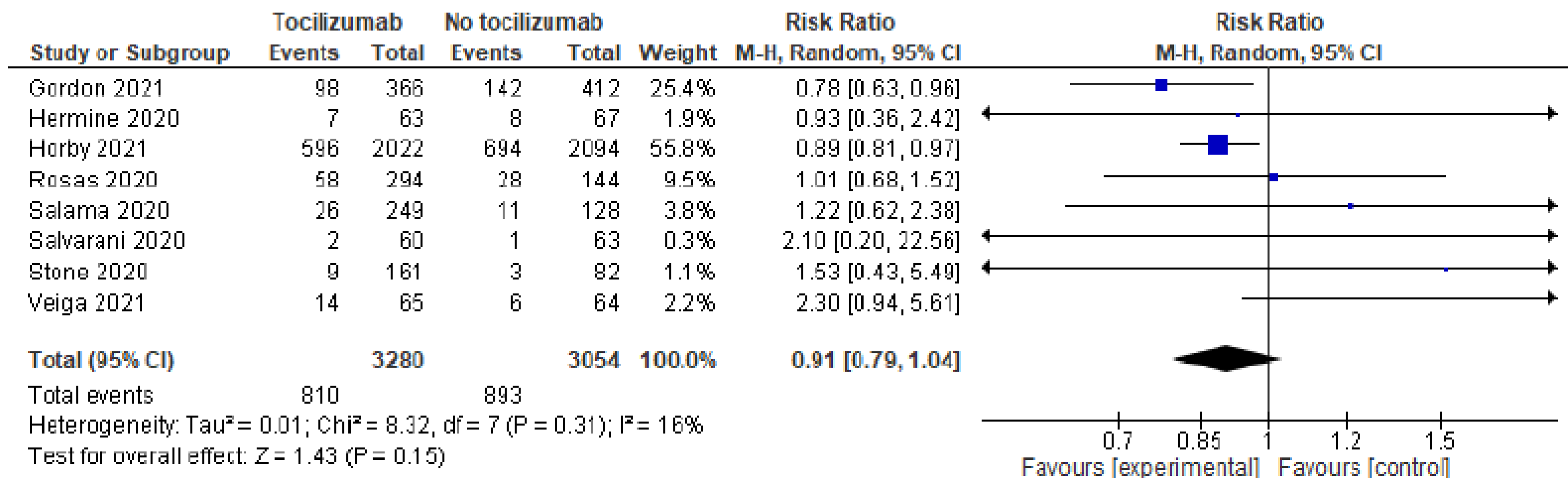








**Figure s6a.** Forest plot for the outcome of mortality for tocilizumab vs. no tocilizumab



Disease Severity	Recommendations for Antiviral or Immunomodulator Therapy	Recommendations for Anticoagulation Therapy
<p><b>Hospitalized but Does Not Require Supplemental Oxygen</b></p>	<p>The Panel <b>recommends against</b> the use of <b>dexamethasone (AIIa)</b> or <b>other corticoster</b></p> <p>There is insufficient evidence to recommend against the routine use of remdesivir. For pat at high risk of disease progression, remdesiv appropriate.</p>	
<p><b>Hospitalized and Requires Supplemental Oxygen</b></p>	<p>Use 1 of the following options:</p> <ul style="list-style-type: none"> <li>• <b>Remdesivir<sup>b,c</sup></b> (e.g., for patients who require minimal supplemental oxygen) <b>(BIIa)</b></li> <li>• <b>Dexamethasone plus remdesivir<sup>b,c</sup></b> <b>(BIIb)</b></li> <li>• <b>Dexamethasone (BI)</b></li> </ul> <p>For patients on dexamethasone with rapidly increasing oxygen needs and systemic inflammation, add a second immunomodulatory drug<sup>d</sup> (e.g., <b>baricitinib<sup>e</sup></b> or <b>tocilizumab<sup>e</sup></b>) <b>(CIIa)</b>.</p>	<p>For nonpregnant patients with D-dimer levels &gt;ULN who are not at increased bleeding risk:<sup>f</sup></p> <ul style="list-style-type: none"> <li>• <b>Therapeutic dose</b> of heparin<sup>g</sup> <b>(CIIa)</b></li> </ul> <p>For other patients:</p> <ul style="list-style-type: none"> <li>• <b>Prophylactic dose</b> of heparin,<sup>g</sup> unless contraindicated <b>(AI)</b></li> </ul>
<p><b>Hospitalized and Requires Oxygen Through a High-Flow Device or NIV</b></p>	<p>Use 1 of the following options:</p> <ul style="list-style-type: none"> <li>• <b>Dexamethasone (AI)</b></li> <li>• <b>Dexamethasone plus remdesivir<sup>b</sup></b> <b>(BIIb)</b></li> </ul> <p>For patients with rapidly increasing oxygen needs and systemic inflammation, add either <b>baricitinib<sup>e</sup></b> <b>(BIIa)</b> or <b>IV tocilizumab<sup>e</sup></b> <b>(BIIa)</b> to 1 of the options above.<sup>d,h</sup></p>	<p>For patients without evidence of VTE:</p> <ul style="list-style-type: none"> <li>• <b>Prophylactic dose</b> of heparin,<sup>g</sup> unless contraindicated <b>(AI)</b></li> </ul>
<p><b>Hospitalized and Requires MV or ECMO</b></p>	<p><b>Dexamethasone<sup>i</sup></b> <b>(AI)</b></p> <p>For patients who are within 24 hours of admission to the ICU:</p> <ul style="list-style-type: none"> <li>• <b>Dexamethasone plus IV tocilizumab (BIIa)</b></li> </ul> <p>If IV tocilizumab is not available or not feasible to use, <b>IV sarilumab</b> can be used <b>(BIIa)</b>.</p>	<p>For patients without evidence of VTE:</p> <ul style="list-style-type: none"> <li>• <b>Prophylactic dose</b> of heparin,<sup>g</sup> unless contraindicated <b>(AI)</b></li> </ul> <p>If patient is started on therapeutic heparin before transfer to the ICU, switch to a <b>prophylactic dose</b> of heparin, unless there is a non-COVID-19 indication <b>(BIII)</b>.</p>

**Figure 2. Therapeutic Management of Adults Hospitalized for COVID-19 Based on Disease Severity**

**Rating of Recommendations:** A = Strong; B = Moderate; C = Optional

**Rating of Evidence:** I = One or more randomized trials without major limitations; IIa = Other randomized trials or subgroup analyses of randomized trials; IIb = Nonrandomized trials or observational cohort studies; III = Expert opinion